BIOINFORMATICS ANALYSIS TOOLS FOR NGS DATA

ANNOTATIONS, VISUALISATION: INTEGRATIVE GENOME BROWSER

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Primary databases

- NCBI GenBank: an annotated collection of all publicly available DNA sequences.
- NCBI RefSeq: A collection of curated, non-redundant genomic DNA, transcript (RNA), and protein sequences produced by NCBI.
- ENA: European Nucleotide Archive: a comprehensive record of the world's nucleotide sequencing information, covering raw sequencing data, sequence assembly information and functional annotation

Commonly used meta databases

- UCSC Genome Bioinformatics: contains the reference sequence and working draft assemblies for a large collection of genomes. It also provides portals to the ENCODE and Neandertal projects
- Ensembl Project by the European Bioinformatics Institute (EBI), European Molecular Biology Laboratory (EMBL), and the Wellcome Trust Sanger Institute (WTSI).

- FlyBase (Drosophila melanogaster)
- Wormbase (Caenorhabditis elegans)
- SGD (Saccharomyces Cerevisiae)
- TAIR (Arabidopsis thaliana)
- Colibri (Escherichia Coli)

And many others – often you'll have to do your research to find the most appropriate resource

Note that each may provide a different user interface, data release methodology, data release update policy etc.

Surprisingly unregulated – and often non-transparent process.

SACCHAROMYCES: GENOME DATABASE

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(The URL is on the course webpage)

http://downloads.yeastgenome.org/curation/chromosomal_feature/saccharomyces_cerevisiae.gff

- Due to a historical limitation (20 years or so ago) and only on Windows → files ended up having a three character extension → .txt, .exe etc.
- This limitation also turned out to be a blessing and it stood the test of time. Makes it easy to see the file type.
- Note: the file extension can be incorrect → mindbogglingly confusing errors may arise then.

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2	pwd			
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4	# get the data, the -o flag stores the result in th	ne sc.gff		
5	curl http://downloads.yeastgenome.org/curation/chro	omosomal_	feature/sacchare	omyc
6				
7	# check to see what files we have here			
8	ls			
9				
10	# check to see how many lines does the file have			
11	wc -l sc.gff			
12				
13	# look at the first ten lines			
14	head sc.gff			
15				
16	# look at the last ten lines			
17	tail sc.gff			
18				
19	<pre># page through the file</pre>			
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- Many common bioinformatics data formats are column based and tab-separated
- Looks like the first format we have to deal with will be the

GFF3 – Generic Feature Format

(search for GFF3 to see the specification for version 3)

http://www.sequenceontology.org/gff3.shtml

Search for GFF3 \rightarrow http://www.sequenceontology.org/gff3.shtml

Tab separated with 9 columns. Missing attributes may be replaced with a dot ightarrow .

- 1. Seqid (usually chromosome, reference point!)
- 2. Source (where is the data coming from)
- 3. Type (usually a term from the sequence ontology)
- 4. Start (interval start relative to the seqid)
- 5. End (interval end relative to the seqid)



- 7. Strand (+/-/.)
- 8. Phase (used to indicate reading frame for coding sequences)
- 9. Attributes (semicolon separated attributes → Name=ABC;ID=1)

Example attribute specification: name=REB1; id=YP33546

These positions

data releases

may change on new

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- Genome browser: one of the most important tools for data analysis
- "Always look at your data", true for statistics as well as for NGS data analysis
- Many genome browsers available: UCSC (web), IGB, Tablet, Artemis, MochiView (ChIP), Chip/SeqMonk (DNA methylation)
- ► Integrative Genomics Viewer
 - Developed by the Broad Institue
 - Awesome and extremely terrible at the same time
 - Works well and is fast enough
 - Data formats: BED, BEDGRAPH, VCF, GTF, BAM, WIG, BIGWIG, FASTA, etc.
 - Usability not perfect

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► Important: the alias file (I'll come back to that later)

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► Save .genome file (contains all the information)

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Custom genome is now part of the genome list

- ► The single most problem with IGV I have encountered so far
- What happens? Sometimes when loading a BAM/BED/VCF/... file, the file appears to be empty.
- Problem: different naming of chromosomes/scaffolds
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- ► Example:

				1. bash				
cont:~	philipp_\$	head /proje	ct/ngs/philip	p/borrelia2	/ref/B31_Schul	zer_refe	rence-goodname:	s.f
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>chr Bo	orrelia bu	rgdorferi B3	1, complete g	enone.				
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CTTCTCC	ATCCCAATI	CCCTAAAGAAGA	TTTAATTAAAAAA	AAAATAAAAAT	ACCCATAA			
TTTACCA	TAATTACAT	AAATTCTATCTT	TTACAATGAAAAT	TATAAATACAT	TGCCTTTA			
TCGGAAT	ATTGACATO	TTATAATGAATG	GATTGAAATACAA	TTTAGCCCCAT	AAATTTTT			
TTACTAT	CCCAACAA	TAAAGATTTTAT	TTCAAATACTTAT	TTCAATTTAGC	TTTCACTA			
TTTACAT	TAC		**********		1. besh			
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cort:~	phi 90_AC	TTGA_C4G2YAND	X_6_20140718	3_20140718_1	.trimmed.bom	cut -f	1.2.3 head	
	HWI-S	T1253F_0165:0	5:2312:17614:	17348#28398_	ACTTGA	16	AE000783.1	
	HWI-S	T1253F_0165:0	5:1213:19639:	35533#20390_	ACTTGA	16	AE000783.1	
	HWI-S	T1253F_0165:0	5:1205:9633:9	2581#20390_4	CTTGA 16	AE000783	3.1	
	HWI-S	T1253F_0165:0	5:1108:4538:8	1682#20390_#	CTTGA Ø	AE000783	3.1	
	HW1-S	12531_0165:0	2104:16451:	10209020390	ACTIGA	16	AE000783.1	
	HW1-S	12535_0165:4	1312:8912:3	9418#20390_F		AE08078:	AE009792 1	
	UNT-S	12535 0165-4	-1105-6746-P	0012#20200	CTTCA A	AC090703	1	
	HWT-S	T1253F @165:0	1108:7441:8	549/20390	CTTGA 0	AE000783	3.1	
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	cort:	- philipp_\$						
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- ► The single most problem with IGV I have encountered so far
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- Problem: different naming of chromosomes/scaffolds
- ► Example:

		1. bash	
cont:~ phi	lipp_\$ head /project/ngs/phi	lipp/borrelia2/ref/B31_S	ichutzer_reference-goodnames.f
asta			
chr Borre	lia burgdorferi B31, complet	e genome.	
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ETTETEGATE	GGAATTCCCTAAAGAAGATTTAATTAAA	AAAAAAATAAAAATAGGCATAA	
TTACCATAA	TTACATAAATTCTATCTTTTACAATGAA	AATTATAAATACATTGCCTTTA	
ICGGAATATT	GACATCTTATAATGAATGGATTGAAATA	CAATTTAGCCCCATAAATTTTT	
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	HWT-ST1253F 0165:6:1108:7441	:85549#20390 ACTTGA 0	AE000783.1
	HWI-ST1253F 0165:6:1202:7530	:80880#20390 ACTTGA 0	AE000783.1
	port:~ philipp \$		

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AE000787.1	lp38
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AE001581.1	cp32-9
AE000785.1	lp25
AE001580.1	cp32-8
AE001579.1	cp32-7
AE001578.1	cp32-6
AE001582.2	lp21
AE001577.1	cp32-4
AE000791.1	cp9
AE001576.1	cp32-3
AE000786.1	lp28-2
AE000794.2	lp28-1
AE000792.1	cp26
AE001575.1	cp32-1
AE000789.1	lp28-4
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	1. bash	
oort:~ phi	lipp_\$ head /project/ngs/philipp/borrelia2/ref/B31_Schu	tzer_reference-goodnames.f
asta		
>chr Borre	lia burgdorferi B31, complete genome.	
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CCCAAATCGA	• • • 1. besh	
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	UNT_ST1253F_0165-6-1108-4538-84682#20308_ACTTCA 0	AC090783 1
	HWT_ST1253E_0165:6:2104:16451:10200#20300_ACTTCA	16 AF909783 1
	HWT-ST1253E @165:6:1312:8912:35418#20390 ACTTGA 0	AF020783.1
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	HWI-ST1253F 0165:6:1105:6746:89913#20390 ACTTGA 0	AE000783.1
	HWI-ST1253F_0165:6:1108:7441:85549#20390_ACTTGA 0	AE000783.1
_	HWI-ST1253F_0165:6:1202:7530:80880#20390_ACTTGA 0	AE000783.1
	cort:~ philipp_\$	

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▶ BAM file has to be sorted and indexed

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▶ BAM file has to be sorted and indexed

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▶ Nothing visible when looking at the full genome

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► Zoom: either use the zoom panel

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4 tracks	chr3.97	108,8	96													15	9M of 513M

> Zoom: or use the mouse to mark region you want to look at



 Max zoom level for showing alignments can be adjusted on the Preference screen



Reads are displayed as red/blue or grey bars



Coverage: how man reads overlap each position of the genome



Comparing two BAM files



Different ranges makes it hard to compare coverage

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► Auto Scale vs. Set Data Range (+log scale)



Auto Scale vs. Set Data Range (+log scale)



Auto Scale vs. Set Data Range (+log scale)



 Mismatches are shown with coloured letters. No letter means match to reference

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Several options for colouring, sorting, grouping



► Example: squished reads



Example: squished reads + no color



► Save current state of IGV



► Save session to file

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► Save session to file

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► Open session



Change settings to relative paths

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6 tracks loaded chr:397,402		67M of 134M

► Change settings to relative paths



Allows to define regions of interest



Allows to define regions of interest



► Click twice to define region



► Click twice to define region



 Region navigator: allows to name regions and jump to their position



 Region navigator: allows to name regions and jump to their position



 Region navigator: allows to name regions and jump to their position



Right click read bar to copy sequence to clipboard



► Export regions to BED file

IGV: DOCUMENTATION



www.broadinstitute.org/software/igv/UserGuide